N-ARACHIDYLANTHRANILIC ACID, A NEW DERIVATIVE FROM ONONIS NATRIX

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ABSTRACT.—From the CHCl₃ extract of the aerial parts of *Ononis natrix*, a new natural product, *N*-arachidylanthranilic acid [1] has been isolated, in addition to the known compounds, gardenin B, xanthomicrol, hymenoxin, 8-hydroxy-6-methoxy-3-undecyl-3,4-dihydroisocoumarin, and medicarpin- β -D-glucoside. The structure of 1 was established by spectroscopic and chemical methods.

The genus Ononis belongs to the family Leguminosae (tribe Trifoleae) and comprises more than 75 annual or perennial species, indigenous to Eurasia, especially the Mediterranean region (1,2). Ononis natrix L. is a perennial herb distributed throughout Jordan (1,3). Infusions of the roots and flowers of O. natrix have been used for the treatment of certain disturbances of the urinary tract and have been reported to have diuretic and antirheumatic properties (4). Compounds isolated from the genus Ononis have also shown antibiotic and molluscicidal activities (5). The genus Ononis is known to produce triterpenoids (2), anthranilic acid derivatives (5), resorcinol derivatives (4-6), dihydroisocoumarins (4,7), a homopterocarpin (7), aromatic lactones (8), flavonoids, and isoflavonoids (5,6). In this paper we describe the isolation and determination of the structure of a new product, namely, N-arachidylanthranilic acid **[1**].

From the CHCl₃ extract of the aerial parts of *Ononis natrix*, five known compounds were isolated, namely, gardenin



B, xanthomicrol, hymenoxin, 8-hydroxy-6-methoxy-3-undecyl-3,4-dihydroisocoumarin, and medicarpin-B-D-glucoside, which were identified by direct comparison with authentic samples and/or with reported physical and spectral data (4, 9-14). In addition, the novel Narachidylanthranilic acid [1] was isolated. This is the first report of the isolation of gardenin B, xanthomicrol, hymenoxin, and medicarpin- β -D-glucose from this genus. Xanthomicrol has shown antispasmodic activity by inhibition of the jejunum in test animals (11), and antimicrobial activity against Aspergillus parasiticus, Candida tropicalis, and Fusarium solani (15). Hymenoxin was found to be cytotoxic to cultured human cells (13). Medicarpin- β -D-glucoside has been previously isolated from the roots of alfalfa (Medicago sativa L.) (14,16), but the ¹³Cnmr data of this substance are reported here for the first time.

The structure of **1** was assigned through interpretation of its spectroscopic properties. The ¹H-nmr spectrum showed signals due to a system of four vicinal aromatic protons at δ 8.77 (dd, $J_{3,4}$ =8 Hz, $J_{3,5}$ =2 Hz, H-3), 8.13 (dd, $J_{6,5}$ =8 Hz, $J_{6,4}$ =2 Hz, H-6), 7.60 (ddd, $J_{4,3}$ =8 Hz, $J_{4,5}$ =7 Hz, $J_{4,6}$ =2 Hz, H-4), 7.11 (ddd, $J_{5,6}$ =8 Hz, $J_{5,4}$ =7 Hz, $J_{5,3}$ =2 Hz, H-5), together with 32 aliphatic protons appearing between δ 1.24–1.42. In addition, two protons appearing as a triplet at δ 2.47 were assigned to those α - to the amide; the two protons which appeared as a quartet at δ 1.77 were assigned to those positioned β - to the amide. The triplet at $\delta 0.88$ (J=7 Hz) was assigned to the terminal methyl group, and a proton of a secondary amide, which appeared at low field as a broad singlet, was observed at δ 10.93 (5.17). The couplings in the ¹H-nmr spectrum of **1** were confirmed by COSY nmr experiments. The methylated derivative 2 showed a similar ¹H-nmr spectrum to that of 1, with the only difference being the singlet signal at δ 3.90 due to the methyl ester of an aromatic acid group. A molecular formula of C₂₇H₄₅O₃N deduced from the ms for **1** was suggested by the molecular ion at m/z 431, and the number of carbons was confirmed by the ¹³C-nmr spectrum with the aid of HETCOR nmr experiments. The base peak of compound 1 appeared at m/z 137, corresponding to free anthranilic acid. This can be interpreted as a result of a McLafferty transposition of an aromatic amide after the loss of an alkylketene from the fragment ion at m/z 179(5,17), and the base peak of the methyl derivative 2 appeared at m/z 151, corresponding to methyl anthranilate. The ir spectrum of 1 showed strong absorptions of the ortho disubstituted benzene ring (1608, 1590, 1455, 755 cm⁻¹), the amide group (3350, 1676, 1535 cm^{-1}), and the aromatic acid (1705, 1418) cm^{-1}) (5,17). Its uv spectrum showed strong absorptions at 253 and 303 nm, consistent with compound 1 being an anthranilic acid derivative (5). With these data and through comparison with those reported for $N-\Delta^{13}$ -docosenoylanthranilic acid (5) and for N-docosenoylanthranilic acid (17), we propose the structure of Narachidylanthranilic acid (N-eicosenoylanthranilic acid) for compound 1.

EXPERIMENTAL

GENERAL EXPERIMENTAL PROCEDURES.—Mps were determined on a Stuart melting-point apparatus and are uncorrected. Ir spectra were determined with KBr pellets on a Jasco IR-810 spectrophotometer. Uv spectra were determined on a Unicon 810 spectrophotometer. ¹H- and ¹³C-nmr spectra were measured on a JEOL GX-270 spectrometer using TMS as internal standard; chemical shifts are reported in δ (ppm) units. Lowresolution ms were recorded on a Varian MAT model CH-5 spectrometer. Si gel was used for cc (Kieselgel 60, Merck) and tlc (Kieselgel 60F₂₅₄).

PLANT MATERIAL.—The aerial parts of Ononis natrix were collected in the vicinity of Dahal, 45 km north of Amman, Jordan, in June 1991. A voucher specimen has been deposited at the Herbarium of the Department of Biological Sciences, Faculty of Science, University of Jordan, Amman, Jordan.

EXTRACTION AND ISOLATION .--- Powdered, air-dried aerial parts of Ononis natrix (17 kg) were repeatedly extracted $(4\times)$ by percolation with EtOH for 10 days (each 20 liters). After the solvent was evaporated, a syrupy residue (916.4 g) was obtained, suspended in H₂O (2 liters), and extracted successively with Et_2O (1 liter \times 3) (fraction A, 180 g), CHCl₃)(1 liter×3)(fraction B, 350 g), and *n*-BuOH (1 liter \times 3) (fraction C, 55 g). Fraction B (350 g) was dissolved in petroleum ether/CHCl₃(2%)(100 ml) and chromatographed over a Si gel (700 g) column (column A) eluted with varying proportions of petroleum ether, CHCl₃, and MeOH mixtures to afford fractions which were collected (100 ml) and combined according to tlc analysis (Me2CO-CHCl2-MeOH, 5:3:2).

Gardenin B.—Elution of the column (column A) with petroleum ether-CHCl₃ (8:2 to 6:4) afforded a solid residue (50 g), which showed two major spots on tlc. The residue was rechromatographed over a Si gel (150 g) column (column B) in petroleum ether-EtOAc (8:2), and the polarity was gradually increased by the addition of EtOAc and EtOAc/MeOH. Gradient elution of column B with petroleum ether-EtOAc (4:6 to 2:8) and EtOAc afforded a solid residue (450 mg) that was recrystallized from MeOH to give gardenin B (300 mg), mp 151°. This compound was identified as 5-hydroxy-6,7,8,4'tetramethoxyflavone by direct comparison (uv, ir, ¹H nmr, ms) with literature data (9,10).

Xanthomicrol.—Continued elution of column B with EtOAc-MeOH (97:3 to 9:1) afforded a residue (4.3 g) which was dissolved in hot MeOH to furnish xanthomicrol (2 g), mp 228°. This isolate was identified as 5,4'-dihydroxy-6,7,8trimethoxyflavone on the basis of spectral and physical data comparison (mp, mmp, uv, ir, ¹H nmr, ms, co-tlc) with an authentic sample (11), and with reported values in the literature (12).

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761

8-Hydroxy-6-methoxy-3-undecyl-3,4-dibydro-

isocoumarin.—Continued elution of column A with petroleum ether-CHCl₃(2:8) and CHCl₃ furnished a residue (45 mg) that, upon crystallization from MeOH, yielded 8-hydroxy-6-methoxy-3-undecyl-3,4-dihydroisocoumarin (28 mg), mp 97°, a known constituent of *O. natrix*, identified by direct comparison with previously reported spectral data (uv, ir, ¹H nmr, ms) (4).

Hymenoxin.—Continued elution of column A with CHCl₃-MeOH (97:3 to 94:6) afforded a solid residue (1.3 g) which, upon treatment with MeOH, yielded hymenoxin (1 g), mp 223–224°. This compound was identified as 5,7-dihydroxy-6,8,3',4'-tetramethoxyflavone on the basis of spectral data comparison (uv, ir, ¹H nmr, ms) with reported values (12,13).

Medicarpin-B-D-glucoside.-Continued elution of column A with CHCl₃-MeOH (91:9 to 85:15) afforded a solid residue (18.3 g), which showed two major spots on tlc. This residue was rechromatographed over Si gel (75 g) (column C) in EtOAc-MeOH (97:3). Elution of the column with EtOAc-MeOH (94:6 to 91:9) afforded a solid residue (56.2 mg) which, upon treatment with MeOH, yielded medicarpin-B-D-glucoside (38 mg), mp 272–273°; ¹³C nmr (CDCl₃, 100 MHz) δ 65.9 (C-2), 40.1 (C-3), 77.7 (C-4), 131.9 (C-5), 110.3 (C-6), 158.4 (C-7), 104.0 (C-8), 156.2 (C-9), 114.1 (C-10), 119.2 (C-1'), 125.1 (C-2'), 106.0 (C-3'), 160.5 (C-4'), 96.3 (C-5'), 160.2 (C-6'), 100.3 (C-1"), 73.1 (C-2"), 77.0 (C-3" or C-5"), 69.6 (C-4"), 76.5 (C-5" or C-3"), 60.6 (C-6"), 55.2 (OMe). This isolate was identified as $(-)-3-\beta$ -Dglucosyl-9-methoxypterocarpan, a known constituent of Medicago sativa roots, by direct comparison with previously reported spectral data (uv, ir, $[\alpha]D, {}^{1}H nmr, ms) (14).$

N-Arachidylanthranilicacid [1].—Continued elution of column C with EtOAc-MeOH (85:15 to 8:2) afforded a solid residue which, on crystallization from MeOH, yielded N-arachidylanthranilic acid [1] (42 mg), mp 86-87°; uv (MeOH) λ max $(\log \epsilon)$ 253 (4.9), 303 (4.6) nm; ir ν max 3350, 1705, 1676, 1608, 1590, 1535, 1475, 1455, 1418, 1282, 1182, 920, 780, 755 cm⁻¹; eims m/z431 (2) (measured 431.3399, calcd 431.3390 for C27H45O3N), 413 (7), 179 (29), 161 (72), 137 (100), 119 (20); ¹H nmr (CDCl₃) δ 0.88 (3H, t, J=7 Hz, H-20'), 1.24–1.42 (32H, m, H-4'–H-19'), 1.77 (2H, q, J=7 Hz, H-3'), 2.47 (2H, t, J=7 Hz, H-2'), 7.11 (1H, ddd, J=8, 7, and 2 Hz, H-5), 7.60(1H, ddd, J=8, 7, and 2 Hz, H-4), 8.13 (1H, dd, J=7 and 2 Hz, H-6), 8.77 (1H, dd, J=8)and 2 Hz, H-3), 10.93 (1H, br s, -NH-); ¹³C nmr (CDCl₃, 100 MHz) & 114.5 (C-1), 142.4 (C-2), 121.0 (C-3), 135.9 (C-4), 123.0 (C-5), 132.0 (C-6), 172.4 (C-1' or COOH), 39.0 (C-2'), 23.0 (C-3'), 25.9-32.3 (C-4' to C-19'), 14.4 (C-20'), 173.2 (COOH or C-1').

Methylation of 1.—A sample of 1 (8 mg) was treated with CH₃I/K₂CO₃ in Me₂CO under reflux for 3 h. The mixture was suspended in H₂O and extracted with EtOAc, the EtOAc solution was evaporated, and the residue was subjected to prep. tlc(n-hexane-EtOAc-MeOH, 8:2:1), to afford compound 2 (5 mg) as an oily liquid. Eims m/z 445 (11), 417 (25), 389 (58), 358 (33), 193 (25), 151 (100), 119 (15); ¹H nmr (CDCl₃) δ 0.88 (3H, t, J=7.2 Hz, H-20'), 1.25–1.43 (32H, m, H-4'–H-19'), 1.77 (2H, q, J=7.2 Hz, H-3'), 2.46 (2H, t, J=7.2 Hz, H-2'), 3.90 (3H, s, OMe), 7.08 (1H, ddd, J=8, 7, and 2 Hz, H-5), 7.55 (1H, ddd, J=8,7, and 2 Hz, H-4), 8.04 (1H, dd, J=7 and 2 Hz, H-6), 8.74 (1H, dd, J=8 and 2 Hz, H-3), 11.05 (1H, br s, -NH-).

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